

BHARATA MATA COLLEGE, THRIKKAKARA

(Affiliated to Mahatma Gandhi University Accredited by NAAC with A+ Grade)

2021- 2024



**" PRODUCTION OF CuSO_4 NANOPARTICLES VIA
COUROPITA GUIANENSIS LEAF EXTRACT FOR
ANTIBACTERIAL AND ANTIOXIDANT PROPERTIES"**

Dissertation submitted to

MAHATMA GANDHI UNIVERSITY, KOTTAYAM

in partial fulfilment of the requirement for the degree of

BACHELOR OF SCIENCE

Submitted by

NANDANA K DIVAKARAN (210021037724)

Under the supervision of

Dr. Sherin Antony

Assistant Professor, Department of Zoology

DEPARTMENT OF ZOOLOGY

BHARATA MATA COLLEGE

THRIKKAKKARA

Date:



CERTIFICATE

This is to certify that the project entitled “ **Production of Nanoparticles Via *Couroupita guianensis* Leaf Extract & its Antimicrobial & Antioxidant Properties**” is a bonafide work done by **NANDANA K DIVAKARAN** with Register No: **210021037724** during 2023-24 in partial fulfilment of the requirement for the award of the Bachelor Degree of Science in Zoology of Mahatma Gandhi University, Kottayam.

Head of the Department

Dr. Simi Joseph P

DECLARATION

I, **NANDANA K DIVAKARAN (210021037724)**, hereby declare that the dissertation work entitled "**Production of CuSO₄ Nanoparticles via *Couroupita guianensis* Leaf Extract for Antimicrobial and Antioxidant Properties**" submitted for the award of a Bachelor's degree in Bharata Mata College, Thrikkakara, is a partial fulfillment of the requirements. This work was done by me during the period from December 2023 to February 2024 under the supervision and guidance of Dr. Sherin Antony, Assistant Professor, Department of Zoology. I affirm that this thesis is original and has not been submitted for any degree, fellowship, or similar qualification by any other candidate to any university.

Date:

Place: Thrikkakara

Signature:

NANDANA K DIVAKARAN

ACKNOWLEDGEMENT

I first give thanks to the All-Powerful God who made it possible for me to finish the thesis on time. It gives me great pleasure to convey my deep appreciation and debt of thanks to Dr. Johnson K M, Principal of Bharata Mata College, Thrikkakara, for giving me a place to complete my project work. I am deeply grateful to Dr. Simi Joseph, Head of the Department of Zoology at Bharata Mata College, Thrikkakara, for her passionate interest, insightful guidance, and critical evaluation of the manuscript. Her unwavering assistance has been extremely helpful in the completion of the thesis. I owe a debt of gratitude to my mentor, Dr. Sherin Antony of the Department of Zoology at Bharata Mata College in Thrikkakara, for all of her support, encouragement, and understanding over the course of my thesis. I sincerely appreciate the time she took to guide throughout the project. I'm quite appreciative of her for helping me with my dissertation. Additionally, I would like to extend my heartfelt appreciation to Dr. Sonia John and Jithin Johnson, Faculty in Charge, Department of Zoology, Bharata Mata College, for their timely support and assistance provided throughout the study, which enabled the successful completion of this project. I sincerely appreciate Mrs. Siji, the Department of Zoology's lab assistant, for her invaluable support and assistance throughout the research project. With profound feelings of appreciation, I express my gratitude to my students and group members for the engaging conversations, assistance, and overall enjoyment we

have shared over the past few months while working together. I am so grateful to my parents and all of my family members, without whose unwavering inspiration, affection, and moral support it would not have been able to finish. Finally, but just as importantly, please accept my sincere thankfulness to God, without whose help this book would never have been published. I apologize to those I haven't spoken to personally and would like to express my gratitude to everyone who helped make this project a success

TABLE OF CONTENTS

Sl .no	CONTENTS	Page No.
1	Abstract	
2	Introduction	
3	Aim and objective	
4	Review of Literature	
5	Materials and methods	
6	Result and discussion	
7	Conclusion	
8	References	

ABSTRACT

Couroupita guianensis, known as the cannonball tree, is a deciduous plant in the Lecythidaceae family. It is famous for its round fruits housed in a tough shell, which gives it its name. The fruit's outer skin starts white and turns blue when exposed to air. Despite being edible, people don't consume it much because of its unpleasant smell, unlike its beautiful blossoms. It has been used for treating various illnesses like malaria and inflammation, although its effectiveness isn't well-documented. Green synthesis, using plant extracts, is an eco-friendly alternative to traditional chemical methods. *C. guianensis* leaves have compounds like flavonoids and terpenoids that offer antiviral, antioxidant, and anti-inflammatory effects. These properties can help with fungal infections, oxidative damage, and microbial issues. The plant may also help combat diseases spread by mosquitoes. Nanoparticles made from the plant have shown potential in medical applications due to their low harm and ability to fight bacteria effectively.

This article discusses to create copper nanoparticles using aqueous extracts derived from *Couroupita guianensis* Aubl. The process is described as eco-friendly, cost-effective, and efficient. Petals, stems, bark, and leaves were utilized to synthesize the nanoparticles. These plant extracts facilitated the reduction of Cu^{2+} ions into CuNP and provided stability. . The newly formed nanoparticles displayed significant antibacterial properties against *Staphylococcus saprophyticus* and *Escherichia coli*. This antibacterial efficacy indicates the potential of utilizing copper nanoparticles, produced inexpensively using leaves of *Couroupita guianensis* Aubl .

Keywords: *couroupita guianensis*, CUS04

INTRODUCTION

Nanoparticles are widely used in various fields such as cosmetics, drug delivery, and pharmaceutical materials to enhance material properties. Nanotoxicology is crucial for safe and sustainable nanotechnology advancement. Nanoscale structures are being organized into superstructures, ensuring nanotechnology's role in multiple technologies. The small size and high surface-to-volume ratio of nanoparticles make them highly appealing. Copper nanoparticles can be synthesized physically, chemically, or biologically, with biosynthesized ones offering improved stability and biocompatibility. Copper nanoparticles are prized for their conductivity, melting point, and cost-effectiveness compared to other metals. They also oxidize to form stable properties. Green nanotechnology is a promising area where functional nanoparticles are produced from iron, zinc, copper, and gold without using harmful chemicals.

The eco-friendly synthesis of nanoparticles is considered an important and cost-efficient process that prioritizes environmental protection. Utilizing natural resources like plant extracts, bacteria, fungi, enzymes, and algae for nanoparticle biosynthesis is a growing field. These resources contain useful compounds that can be used to create nanoparticles, offering advantages like energy efficiency, cost-effectiveness, and environmentally safe chemicals. Nanoparticles produced through biosynthesis have shown promising medicinal properties, including antimicrobial, antioxidant, antimalarial, anti-inflammatory, anti-diabetic, and anticancer effects. In this study,

active components from *C. guianensis* extract were used to produce copper nanostructures for antimicrobial purposes. The advancement and use of nanotechnologies have led to numerous possibilities in research and industry, creating new materials with a wide range of applications across various sectors. Nanoparticles, as a type of nanomaterial, have garnered considerable interest for their diverse uses in different fields.



Image of *Couroupita guianensis* Aubl flower

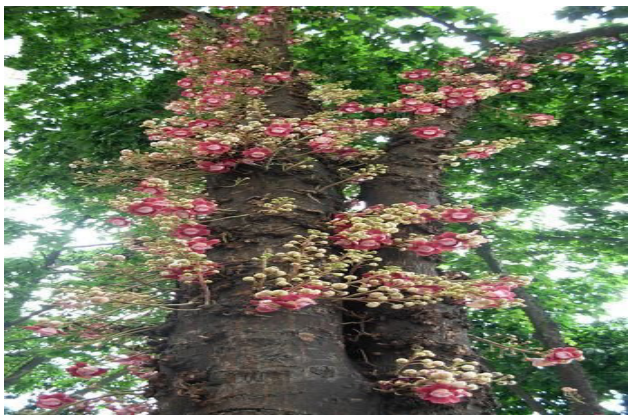


image of *Couroupita guianensis* tree is known by a variety of common names including cannonball tree.

AIM AND OBJECTIVE

AIM

This study aims to use leaf extract from *Couroupita guianensis* to create copper sulfate nanoparticles and assess their antibacterial and antioxidant qualities.

OBJECTIVE

To Evaluate the effectiveness of the copper sulphate nanoparticles against bacteria. Examine the potential antioxidant properties of the plant extract.

REVIEW OF LITERATURE

The Lecythidaceae family comprises large tropical deciduous trees like the cannonball tree (*Couroupita guianensis* Aubl.). The active compounds found in various parts of the cannonball tree, such as leaves, stems, flowers, and bark, have potential healing properties. This potential is attributed to essential oils, glycosides, ketosteroids, isatin, indurubin, and phenolic chemicals present in the plant. The chemical components of the tree's leaves include hydroxycinnamic acids, rosmarinic acid, and various other substances. Leaf extracts of the cannonball tree show antibacterial and antifungal properties that can help in treating various ailments). The Lecythidaceae family consists of large tropical deciduous trees like the cannonball tree (*Couroupita guianensis* Aubl.). The medicinal properties of various parts of the plant, such as leaves, stems, flowers, bark, etc., contain bioactive compounds that can be used for healing purposes. These healing properties are attributed to essential oils, glycosides, ketosteroids, isatin, indurubin, and phenolic chemicals found in the plant (Pandurangan et al., 2018).

The leaf extracts of *C. guianensis* contain phenolic and volatile compounds that exhibit antibacterial and antifungal effects, making them effective in treating various diseases (Elumalai et al., 2012).

Because EO water uses a small amount of salt solution and does

not contain extra chemical additives, it has a reduced environmental impact in terms of chemicals (Kim et al., 2000).

Nanoparticles are now widely used in various aspects of our lives, including cosmetics and drug delivery systems. They play a significant role by improving the properties of various materials in therapeutic, biosensor, and pharmaceutical applications (Nithiyavathi et al., 2021, Poovendran et al., 2020, George et al., 2022). Nanoparticles have generated significant interest due to their extremely small size and large surface-to-volume ratio (Theophil Anand et al., 2019, Manjula et al., 2018).

Copper nanoparticles were produced through physical, chemical, and biological methods (EL-Din Hassan et al., 2018).

Copper nanoparticles are increasingly popular because of their high electrical conductivity, elevated melting point, minimal electrochemical migration, and cost-effectiveness compared to metals like silver, gold, platinum, and palladium (Rajesh et al., 2018, Rajeshkumar and Rinitha, 2018, Rehana et al., 2017).

Moreover, copper nanoparticles have the advantage of oxidizing to form stable nanoparticles with consistent chemical and physical properties. The field of green nanotechnology has emerged, focusing on the creation of functional nanoparticles using iron, zinc, copper, and gold without the need for toxic chemicals (Nazar, 2018).

The green synthesis of nanoparticles (NPs) is considered a safer, more cost-effective, and environmentally friendly approach compared to physical and chemical methods [1].(Abdelghany, T.M.; Al-Rajhi, A.M.H.; al Abboud, M.A.; Alawlaqi, M.M.; Ganash Magdah, A.; Helmy, E.A.M.; Mabrouk, A.S.)

Metal NPs like Au, Ag, Zn, and Cu have been successfully synthesized and studied for various medical purposes. Among these, silver (Ag) NPs are commonly produced using plant extracts. Ag NPs are renowned for their distinct features such as excellent thermal and electrical conductivities, enhanced stability, and promising biological effects [2].(Netala, V.R.; Kotakadi, V.S.; Nagam, V.; Bobbu, P.; Ghosh, S.B.; Tartte, V.)

Biologically synthesized Ag NPs are gaining significant attention in biomedical fields, particularly in drug development for infectious diseases, drug delivery systems, diagnostics, and mosquito control [3].(Huy, T.Q.; Huyen, P.T.M.; Le, A.-T.; Tonzzer, M)

Ag NPs ranging from 1 to 100 nm in size, coupled with a large surface area and highly reactive surfaces, are attracting significant interest [4,5,6,7,8].(Elangovan, K.; Elumalai, D.; Anupriya, S.; Shenbhagaraman, R.; Kaleena, P.K.; Murugesan, K.4.Salata, O.5.Shanmugasundaram, T.; Balagurunathan, R.6."Benelli, G.; Govindarajan, M."7."Benelli, G.; Govindarajan, M.8)

In this research, extracts from *Lagerstroemia speciosa* (L. *speciosa*) fruits and *Couroupita guianensis* (C. *guianensis*)

flowers were utilized in the creation of Ag NPs. The plant *L. speciosa*, commonly known as "Jarul," belongs to the Lythraceae family [9].("Sondi, I.; Salopek-Sondi, B.)

These plant parts contain various phytochemicals like alkaloids, terpenoids, flavonoids, and others [10,11].("Myint, P.P.; Soe, M.T.; Hlaing, H.H", "Pareek, A.; Suthar, M.; Rathore, G.S.; Bansal, V")

Moreover, both the leaf and flower extracts from *L. speciosa* have been extensively examined for the green synthesis of Ag NPs [8] and studied for a range of biological activities, including antibacterial, anti-diabetic, and anti-inflammatory properties [12,13].("Al-Snafi, A.E. ", "Sharmin, T.; Rahman, M.; Mohammadi, H.")

Additionally, components of *C. guianensis* (also known as ayahuma and cannonball tree), a plant from the Lecythidaceae family, have been explored for their potential in anticancer, antifungal, and anti-inflammatory applications. Extracts from these plant parts have been traditionally used to alleviate common cold symptoms, stomach aches, and malaria [14,15].(: "Sumathi, S.; Anuradha, R, "Sheba, L.A.; Anuradha, V.")

Recent findings have demonstrated that leaf and fruit extracts of *C. guianensis* are effective in the rapid and cost-effective production of Ag NPs and controlling the dengue vector *Aedes aegypti* [16].

Yet, there has been limited research on the biological properties of Ag NPs synthesized from plant extracts. Therefore, this study aims to investigate the *in vitro* antibacterial, antioxidant, larvicidal, and cytotoxic properties of Ag NPs produced using extracts from

L. speciosa fruits and *C. guianensis* flowers. The combination of bioactive compounds from these sources can lead to Ag NPs with diverse biological effects.

MATERIAL AND METHODS

1. Collection of plants and substances

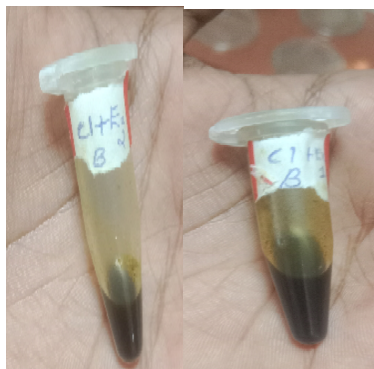
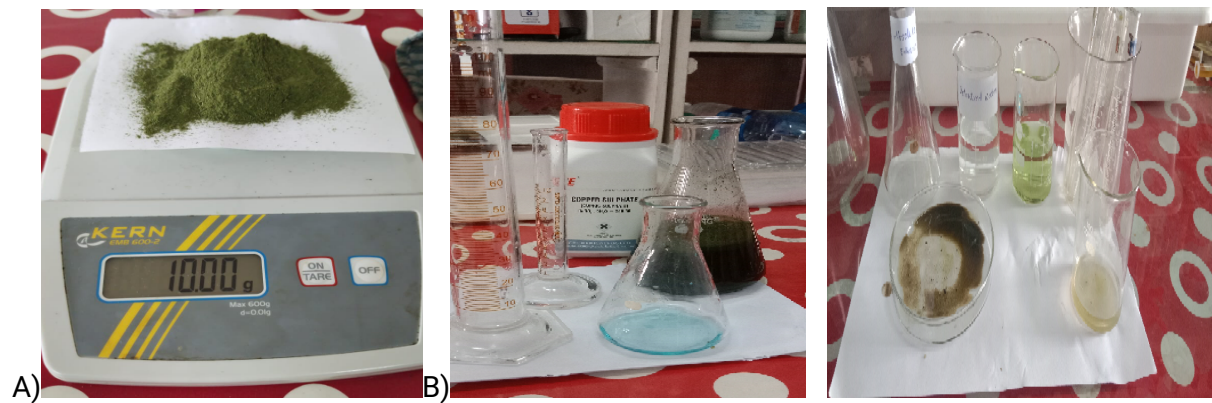
Fresh and healthy leaves, of *C. guianensis* were gathered from the college premises. They were cleaned, dried in the shade for 5-7 days, and then ground into a fine powder using an electric mixer

2. Preparation of plant extracts

To make the plant extract, 10 grams of powdered leaves, stems, petals, and barks were combined with 100 ml of deionized water and boiled at 60 °C for 30 minutes. The mixture was then filtered using Whatman No.1 paper to eliminate any residue and stored at 4 °C for future use.

3.To synthesize CuNPs, 0.25 g of CuSO₄ was dissolved in 50 ml of water. Then, 4 ml of *C. guianensis* extract was added slowly while stirring for 10 minutes until a dark green color appeared. The mixture was stirred at 60 °C for 1-2 hours, left at room temperature for 12-14 hours, and then centrifuged at 10,000 rpm for 10 minutes. The resulting solid was washed with water and ethanol several times to remove impurities before drying in an oven at 70 °C for 10 hours.

PREPARATIONS OF NANOPARTICLES



RESULT AND DISCUSSION

The compound was tested for its ability to inhibit the growth of two bacterial strains, *Staphylococcus aureus* and *Escherichia coli*. The results showed varying levels of antimicrobial activity, ranging from moderate to good. The antibacterial activity was assessed using the agar well diffusion technique. The diameter of the inhibition zone was measured after 24 hours of bacterial incubation at 30 °C. A filter paper disc saturated with a sample of the compound was placed on agar medium seeded with the test organism. The clear zone of inhibition surrounding the sample was measured to determine its inhibitory power. The antimicrobial activity of the tested compounds was evaluated against both gram-positive and gram-negative bacteria.

Many studies have shown that nanomaterials have unique characteristics that make them suitable for use in personal care products, cosmetics, medicines, drug delivery systems, and textiles. Green synthesis of nanoparticles offers a more sustainable alternative to traditional methods that involve toxic solvents, chemicals, and harsh reaction conditions like high temperatures and pressure. This environmentally friendly approach allows for the cost-effective and efficient production of nanoparticles with improved stability, effectiveness, purity, and uniformity in size.

E. coli, or *Escherichia coli*, is a bacteria commonly found in the intestines of humans and animals. While most are harmless, some strains can lead to food poisoning. *E. coli* is extensively studied in microbiology and genetics due to its quick growth and well-known genetics. Although it can be treated with antibiotics like penicillin, cephalosporins, fluoroquinolones, and aminoglycosides, misuse has led to the rise of antibiotic-resistant strains, presenting challenges in treating infections caused by *E. coli*.

Antibacterial disc	3.6cm
Ethanol	0.5cm

DMSO	0.4cm
Ethanol + Nanoparticles (100µl)	1.1cm
DMSO + Nanoparticles (100µl)	1.6cm

a) Antibacterial effect shown against E.coli

Staphylococcus saprophyticus is a bacteria commonly found in the gastrointestinal and genital tracts of humans and is a frequent cause of UTIs, especially in young sexually active females. It is typically treatable with antibiotics like trimethoprim-sulfamethoxazole, nitrofurantoin, and fluoroquinolones, but resistance can develop. This resistance can present difficulties in effectively treating infections caused by this bacterium.

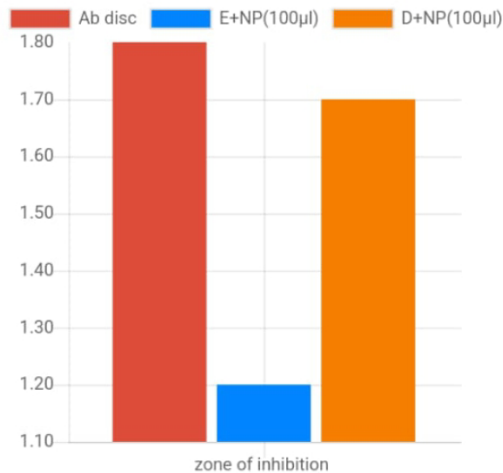
Antibacterial disc	1.8cm
Ethanol	0.3cm
DMSO	0.2 cm
Ethanol +Nanoparticles (100µl)	1.2cm
DMSO+Nanoparticles (100µl)	1.7cm

(b) Antibacterial effect shown against Staphylococcus saprophyticus

The study focused on creating CuO nanoparticles using Couroupita guianensis, also known as the cannonball tree, which is an interesting topic of research. Green synthesis techniques are becoming popular due to their eco-friendly nature and potential uses in different fields. CuO nanoparticles made with Couroupita guianensis have shown promising antibacterial and antifungal properties. They have proven effective against a variety of microorganisms, including Escherichia coli, Staphylococcus saprophyticus, and Candida albicans. The CuO nanoparticles also have antioxidant properties, with studies showing they have significant antioxidant activity by scavenging free radicals and preventing oxidative damage. This makes

them valuable for potential applications in medicine and cosmetics, where antioxidants are used for protection against diseases and aging.

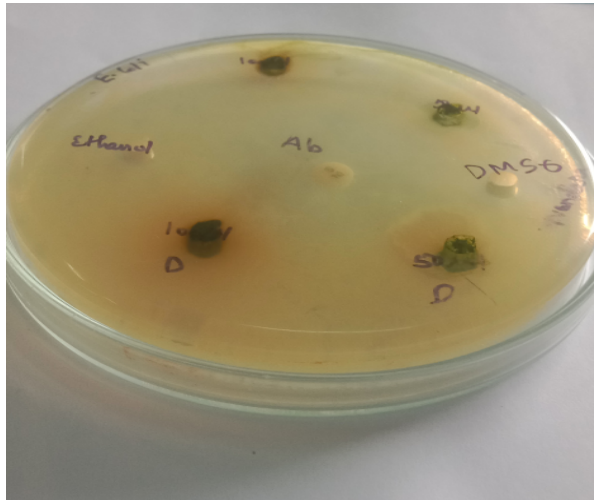
Bar graph showing zone of inhibition against *Staphylococcus saprophyticus*



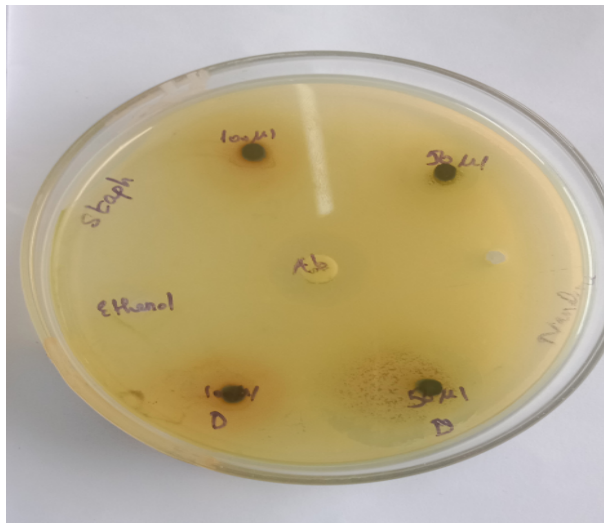
ANTIBACTERIAL ACTIVITY

The tested compound was prepared and evaluated for its antibacterial activity against *Staphylococcus aureus* and *Escherichia coli* through the diffusion plate method. Results showed a range of moderate to good antimicrobial activity. The bacteria were cultured on Czapek-Dox agar medium and the inhibition zones were measured after 24 hours.

of incubation at 30°C. The diameter of the clear zones surrounding the sample was used to determine the inhibitory power of the compound. Various references were consulted for the examination of the antimicrobial activity against the tested bacteria.



Antibacterial effect shown against *Escherichia coli*



Antibacterial effect shown against *Staphylococcus aureus*

CONCLUSION

In the present work, the synthesis of copper nanoparticles was investigated using aqueous extracts of *Couroupita Guianensis* Aubl leaves as a stabilizer agent. A simple and feasible method is presented in this work to produce copper nanoparticles with

desirable functional properties. The characterization of CuNPs were subjected to the antibacterial activity for gram-negative and gram-positive bacterial strains, and exhibited good results for *E. coli* and *Staphylococcus saprophyticus*. The CuNPs may be an effective product used for many biomedical applications, which could be a potential agent.

REFERENCES

1. S. Logambal a, C. Maheswari b, S. Chandrasekar b, T. Thilagavathi c, C. Inmozhi d, S. Panimalar e, F.A. Bassyouni f, R. Uthrakumar a, Mohamed Ragab Abdel Gawwad g, Reem M. Aljowaie h, Dunia A. Al Farraj h, K. Kanimozhi "Synthesis and characterizations of CuO nanoparticles using *Couroupita guianensis* extract for and antimicrobial applications" Volume 34, Issue 3 April 2022.
2. Anand et al., 2021. G.T. Anand, S.J. Sundaram, K. Kanimozhi, R. Nithiyavathi, K. Kaviyarasu. Microwave assisted green synthesis of CuO nanoparticles for environmental applications Mater. Today: Proc., 36 (2021), pp. 427-434
3. Anna Sheba and Anuradha, 2020 An updated review on *Couroupita guianensis* Aubl: a sacred plant of India with myriad medicinal properties J. Herbmed. Pharmacol., 9 (2020), pp. 1-11
4. Venkatadri Babu 1,†ORCID, Selvaraj Arokiyaraj 2,*†, Swathi Pon Sakthi Sri 3, Mary George 3, Rameshkumar Marimuthu Ragavan 4, Dinesh Dharmalingam 5, Taehwan Oh 6, Subramaniyan Ramasundaram 6,* and Paul Agastian 1,* "Antibacterial, Antioxidant, Larvicidal and Anticancer Activities of Silver Nanoparticles Synthesized Using Extracts from Fruits of *Lagerstroemia speciosa* and Flowers of *Couroupita guianensis*" 12 November 2022 Molecules 2022, 27(22), 7792;
5. Abdelghany, T.M.; Al-Rajhi, A.M.H.; al Abboud, M.A.; Alawlaqi, M.M.; Ganash Magdah, A.; Helmy, E.A.M.; Mabrouk, A.S. Recent Advances in Green Synthesis of Silver Nanoparticles and Their Applications: About Future Directions. A Review. Bionanoscience 2018, 8, 5–16. [Google Scholar] [CrossRef]
6. Jerry O. Adeyemi, 1,* Ayodeji O. Oriola, 1 Damian C. Onwudiwe, 2,3 and Adebola O. Oyedeji 1 "Plant Extracts Mediated Metal-Based Nanoparticles: Synthesis and Biological Applications" 2022 May; 12(5): 627. Published online 2022 Apr 24.
7. Suresh Chand Mali, Shani Raj, Rohini Trivedi "Biosynthesis of copper oxide nanoparticles using *Enicostemma axillare* (Lam.) leaf extract" Volume 20 December 2019
8. DR. MAHIPAL SINGH SHEKHAWAT "Investigations on vitro regeneration of *Couroupita guianensis* Aubl (Nagalingam tree) - A threatened but medicinally important plant" May 2014
9. G.T. Anand, S.J. Sundaram, K. Kanimozhi, R. Nithiyavathi, K. Kaviyarasu Microwave assisted green synthesis of CuO nanoparticles for environmental applications Mater. Today: Proc., 36 (2021), pp. 427-434

- 10.K. Kaviyarasu, C.M. Magdalane, K. Anand, E. Manikandan, M. Maaza Synthesis and characterization studies of MgO: CuO nanocrystals by wet-chemical method Spectrochim. Acta Part A Mol. Biomol. Spectrosc., 142 (2015), pp. 405-409
- 11.Khan MR, Kihara M, Omoloso AD. Antibiotic activity of *Couroupita guianensis*. J Herbs Spices Med Plants. 2003;10(3):95-108.
- 12.Pinheiro MM, Fernandes SB, Fingolo CE, Boylan F, Fernandes PD. Anti-inflammatory activity of ethanol extract and fractions from *Couroupita guianensis* Aublet leaves. J Ethnopharmacol. 2013;146(1):324-30. doi: 10.1016/j.jep.2012.12.053
- 13.Uppala PK, Murali Krishna B, Atchuta Kumar K, Vinay Ramji DJ. Evaluation of anti-coagulant activity of the chloroform and aqueous extracts of the leaves of *Couroupita guianensis*. Int J Pharm Pharm Res. 2016;6(4):189-99
- 14.Wang Y., Wang H., Khan M.S., Husain F.M., Ahmad S., Bian L. Bioconjugation of Gold Nanoparticles with Aminoguanidine as a Potential Inhibitor of Non-Enzymatic Glycation Reaction. J. Biomol. Struct. Dyn. 2021;39:2014–2020. doi: 10.1080/07391102.2020.1749131. [PubMed] [CrossRef] [Google Scholar]
15. Ashraf J.M., Ansari M.A., Fatma S., Abdullah S.M.S., Iqbal J., Madkhali A., Hamali A.H., Ahmad S., Jerah A., Echeverria V., et al. Inhibiting Effect of Zinc Oxide Nanoparticles on Advanced Glycation Products and Oxidative Modifications: A Potential Tool to Counteract Oxidative Stress in Neurodegenerative Diseases. Mol. Neurobiol. 2018;55:7438–7452. doi: 10.1007/s12035-018-0935-x. [PubMed] [CrossRef] [Google Scholar]
16. Gahlawat G., Choudhury A.R. A Review on the Biosynthesis of Metal and Metal Salt Nanoparticles by Microbes. RSC Adv. 2019;9:12944–12967. doi: 10.1039/C8RA10483B. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
17. Ovais M., Khalil A.T., Islam N.U., Ahmad I., Ayaz M., Saravanan M., Shinwari Z.K., Mukherjee S. Role of Plant Phytochemicals and Microbial Enzymes in Biosynthesis of Metallic Nanoparticles. Appl. Microbiol. Biotechnol. 2018;102:6799–6814. doi: 10.1007/s00253-018-9146-7. [PubMed] [CrossRef] [Google Scholar]

18. Watt J., Cheong S., Tilley R.D. How to Control the Shape of Metal Nanostructures in Organic Solution Phase Synthesis for Plasmonics and Catalysis. *Nano Today*. 2013;8:198–215. doi: 10.1016/j.nantod.2013.03.001. [CrossRef] [Google Scholar]
19. Kandi V., Kandi S. Antimicrobial Properties of Nanomolecules: Potential Candidates as Antibiotics in the Era of Multi-Drug Resistance. *Epidemiol. Health*. 2015;37:e2015020. doi: 10.4178/epih/e2015020. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
20. Wang Y., Xia Y. Bottom-up and Top-down Approaches to the Synthesis of Monodispersed Spherical Colloids of Low Melting-Point Metals. *Nano Lett*. 2004;4:2047–2050. doi: 10.1021/nl048689j. [CrossRef] [Google Scholar]
21. Mittal A.K., Chisti Y., Banerjee U.C. Synthesis of Metallic Nanoparticles Using Plant Extracts. *Biotechnol. Adv.* 2013;31:346–356. doi: 10.1016/j.biotechadv.2013.01.003. [PubMed] [CrossRef] [Google Scholar]
22. Cao G. *Nanostructures and Nanomaterials*. Imperial College Press; London, UK: 2004. [Google Scholar]
23. Singh J., Dutta T., Kim K.-H., Rawat M., Samddar P., Kumar P. “Green” Synthesis of Metals and Their Oxide Nanoparticles: Applications for Environmental Remediation. *J. Nanobiotechnol.* 2018;16:84. doi: 10.1186/s12951-018-0408-4. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
24. Das R.K., Pachapur V.L., Lonappan L., Naghdi M., Pulicharla R., Maiti S., Cledon M., Dalila L.M.A., Sarma S.J., Brar S.K. Biological Synthesis of Metallic Nanoparticles: Plants, Animals and Microbial Aspects. *Nanotechnol. Environ. Eng.* 2017;2:18. doi: 10.1007/s41204-017-0029-4. [CrossRef] [Google Scholar]
25. Husen A. *Nanoscience and Plant–soil Systems*. Springer; Cham, Switzerland: Berlin, Germany: 2017. *Gold Nanoparticles from Plant System*:

Synthesis, Characterization and Their Application; pp. 455–479. [CrossRef] [Google Scholar]

26. Ankamwar B. Biosynthesis of Gold Nanoparticles (Green-Gold) Using Leaf Extract of Terminalia Catappa. *E-J. Chem.* 2010;7:1334–1339. doi: 10.1155/2010/745120. [CrossRef] [Google Scholar]

27. Adeyemi J.O., Elemike E.E., Onwudiwe D.C. ZnO Nanoparticles Mediated by Aqueous Extracts of Dovyalis Caffra Fruits and the Photocatalytic Evaluations. *Mater. Res. Express.* 2019;6:125091. doi: 10.1088/2053-1591/ab5bcb. [CrossRef] [Google Scholar]

28. Li X., Xu H., Chen Z.-S., Chen G. Biosynthesis of Nanoparticles by Microorganisms and Their Applications. *J. Nanomater.* 2011;2011:270974. doi: 10.1155/2011/270974. [CrossRef] [Google Scholar]

29. Zhang X.-F., Liu Z.-G., Shen W., Gurunathan S. Silver Nanoparticles: Synthesis, Characterization, Properties, Applications, and Therapeutic Approaches. *Int. J. Mol. Sci.* 2016;17:1534. doi: 10.3390/ijms17091534. [PMC free article] [PubMed] [CrossRef] [Google Scholar]